Original Investigation

Serum Heat Shock Protein Levels and the Relationship of Heat Shock Proteins with Various Parameters in Chronic Obstructive Pulmonary Disease Patients

Ramazan Ünver¹, Figen Deveci¹, Gamze Kırkıl¹, Selda Telo², Dilara Kaman², Mutlu Kuluöztürk^ı

1 Department of Chest Diseases, Fırat University Faculty of Medicine, Elazığ, Turkey

2 Department of Biochemestry, Fırat University Faculty of Medicine, Elazığ, Turkey

Abstract **OBJECTIVES:** Chronic Obstructive Pulmonary Disease (COPD) is accompanied by increased cellular stress and inflammation. Most of the Heat Shock Proteins (HSPs) have strong cytoprotective effects. The role of HSPs in COPD pathogenesis has not determined completely. We investigated the serum level of HSPs in COPD patients, smokers without COPD and healthy non-smoking controls. Also, we evaluated the relationship of HSPs with various parameters (inflammatory, oxidative, functional status, quality of life) in COPD patients.

MATERIAL AND METHODS: The levels of stress protein (HSP27, HSP70, HSP60, HSP90, CyPA), interleukin-6, C-reactive protein and malondialdehyde were measured in 16 healthy non-smoker, 14 smokers without COPD and 50 patients with stable COPD. Pulmonary function tests (PFT) and arterial blood gases parameters were measured. Health Related Quality of Life was evaluated and exercise capacity was measured with 6 minute walking test.

RESULTS: Only HSP27 levels was significantly higher in COPD patients when compared with both healthy non-smoker and smokers without COPD (for both, p< 0.001). There was a weak-moderate negative correlation between serum levels of HSP27 and PFT parameters and between HSP27 levels and PaO₂. Serum levels of HSP27 showed a weak-moderate positive correlation with symptom, activity and total scores. Subjects evaluated only smokers without COPD and patients with COPD; HSP27 had an area under the curve (AUC) in the receiver operating characteristic (ROC) curve of 0.819 (0.702-0.935; 95% CI; p= 0.000).

CONCLUSION: Increased serum levels of HSP27 was found in COPD patients and our results showed sensitivity and specificity of serum HSP27 as diagnostic markers for COPD.

KEYWORDS: COPD, heat shock protein, oxidative stress, hypoxia *Received: 22.12.2015 Accepted: 13.03.2016*

INTRODUCTION

There is prominent inflammatory response and oxidant-antioxidant imbalance in Choronic Obstructive Pulmonary Disease (COPD). Also, persisting inflammatory reactions continue in COPD patients despite cessation of smoking. Cigarette smoking is the major risk factor for COPD. Cigarette smoke contains multipl free radicals and these toxic substances are believed to induce an inflammatory response by adversely affecting oxidant/antioxidant and protease/ anti-protease balance in the lung. In fact, COPD don't develop all smokers. The majority of long-term smokers do not develop COPD suggests that failure of compensatory mechanisms that protect the lung from reactive oxygen species (ROS) or xenobiotic materials contributes to development of the disease. The expression of antioxidant genes believed to be important in protection of the lung from cigarette smoke-induced injury. Recent studies indicate that a complex molecular cascade termed the ''unfolded protein response'' (UPR) plays an important role in the regulation of expression of a variety of antioxidant, xenobiotic metabolizing and pro- and anti-inflammatory genes [1].

Heat shock proteins (HSPs) are chaperones that catalyze the proper folding of nascent proteins and the refolding of denatured proteins. HSPs have a role either the renaturation or the destruction of damaged proteins under stressful conditions such as heat, bacterial or viral infections [2]. HSP27 was first reported to contribute to heat shock resistance; subsequently, its involvement in diverse protective mechanisms against toxicity mediated by aberrantly folded proteins or oxidative-inflammatory conditions has also been confirmed [3]. Under normal physiological conditions the synthesis of most HSPs is low. However, when organisms endure stress such as heat shock and inflammation, where protein damage is increased, certain HSP are induced and expressed at high levels [4]. Increased HSPs levels showed in COPD patients. HSPs, especially HSP60 may have a role in COPD pathogenesis and some HSPs might be used as possible

serum markers for determining COPD in the smoking subjects [5,6]. To our knowledge, the role of HSPs in pathogenesis and diagnosis of COPD has been investigated in few studies.

The aim of our study was to investigate whether the serum levels of various HSPs are elevated in smokers without COPD and COPD patients and to determine the relationship between HSPs and several parameters (inflammatory, oxidative, functional status and quality of life) in COPD patients.

PATIENTS and METHODS

Subjects

This study was done between September 2012 and April 2013. This case control study included 80 patients with COPD and controls. Healthy non-smoker volunteers (n= 16), smokers without COPD ($n= 14$), patients with COPD ($n= 50$) were evaluated.

Control group, consisted of 16 healthy non-smoking subjects and 14 smokers without COPD, had normal pulmonary function parameters and they had not any lung disease. All subjects were selected with Stratified Random Sampling Method from amongst the hospital staff. The age and sex of the control subjects were similar to COPD patients.

Fifty stable COPD patients enrolled into the study and they were taken from a hospital respiratory out-patient clinic. COPD was diagnosed according to Global Initiative for Chronic Obstructive Lung Disease (GOLD) criteria [7]. In addition, the classification of airflow limitation severity was evaluated according to GOLD guidelines [7]. Patients with no evidence of an exacerbation for one month before study were accepted as clinically stable. Acute exacerbation as defined by GOLD, use of systemic steroids within the past 14 days, asthma, autoimmune diseases, lung cancer, known 1-antitrypsin deficiency and known cardiopulmonary co-morbidity were considered as exclusion criteria.

Ethical approval was obtained by the institutional review board (31.05.2012-09) and informed consent was obtained from each subject.

Age, gender and smoking history were asked and the body mass index (BMI) and pulmonary function tests (PFTs) was detected in all subjects. Six minute walking test (mwt), arterial blood gases (ABG) analysis were done in COPD patients and Health Related Quality of Life (HRQL) also evaluated in COPD patients.

Pulmonary Function Testing

The pulmonary function tests were done using a spirometry device (Ultima CPX 790705-205, St. Paul, MN, USA). The standard spirometric examination was conducted according to European Respiratory Society (ERS) criteria [8]. Forced expiratory volume in 1 s (FEV₁) and forced vital capacity (FVC) are expressed as percentages of predicted values (FEV₁% pred and FVC% pred) according to the prediction equations of the ERS [8].

HRQL was assessed in COPD patients using the Turkish version of St. Georges Respiratory Questionnaire (SGRQ) [9,10]. The questionnaire was applied to COPD patients by the same interviewers. The SGRQ has been used extensively for assessing quality of life in patients with COPD and several other chronic lung diseases [11]. It contains 50 items with 76 weighted responses that cover three domains: symptomsdistress due to respiratory symptoms, activity-disturbances of physical activity and impact-overall impact on daily life and well-being. In addition to the domain scores, there is also a total score [9]. The SGRQ is scaled from zero to 100 (with zero representing the best health-related quality of life).

Exercise Performance

Exercise performance was evaluated by the 6 mwt according to the American Thoracic Society Guideline [12].

Arterial blood gas measurement

Arterial blood gas samples of COPD patients were taken at rest, in a sitting position and in room air at the room temperature. Samples were measured by a blood gas analyse device (Rapid lab 348. Biobak., Chiron, Bayer Diagnostic, UK).

Measurement of serum HSPs, CRP, IL-6, CRP and MDA levels

Blood samples were collected between at 8.30-9.30 following 10-hours starvation. Serum was acquired after centrifugation and aliquots were kept frozen at -20°C until further testing. HSP27, HSP70, HSP60, HSP90, CyPA and interleukin-6 (IL-6) were determined using adapted enzyme-linked immunosorbent assay (ELISA) kits according to kits protocol. Levels of HSP27, HSP60, HSP70, HSP90, CyPA, and IL-6 were determined using adapted ELISA kits [(Boster Biological technology., Ltd. (Catalog no: EK0881), assaypro (Catalog no: EH5505-1), Hangzhou eastbiopharm co. ltd. (Catalog no: CK-E11197), Hangzhou eastbiopharm co. ltd. (Catalog no: CK-E11190), Hangzhou eastbiopharm co. ltd. (Catalog no: CK-E90142), Boster immumoleader (Catalog no: EK0410), respectively] according to kits protocol.

The concentration of serum malondialdehyde (MDA) was determined by High-performance liquid chromatography (HPLC) using Immuchrom commercial kit (ImmuChrom GmbH, Munich, Germany) according to kit protocol.

Serum levels of C-reactive protein (CRP) were routinely analyzed by the Central Laboratory at the hospital.

Statistics

Data were analyzed using the statistical package for the social sciences (SPSS) software statistical program. Results were given as median and 95% CI. A p value of < 0.05 was considered statistically significant. Statistical analysis was performed using Kruskal-Wallis test for multiple-group comparisons; Mann-Whitney U test was performed to test any observed differences for significance and results were interpreted according to Benferroni correction. Chi-square test was performed to compare gender distribution between groups. Spearman's correlation was used to assess nonparametric data. Receiver operating characteristic (ROC) curves were plotted to show sensitivity and specificity of the evaluated HSPs.

RESULTS

Age, gender and BMI were found similar between the patient population and control subjects (p> 0.05). Patient characteristics and PFT parameters were shown in Table 1.

There was no statistically significant difference in the levels of HSP70, HSP90, HSP60 and CyPA between groups (p> 0.05) (Table 2). The serum levels of HSP27 were statistically higher in COPD patients than in both healthy non-smoker and smokers without COPD (for both p< 0.001) (Table 2, Figure 1A). There was no statistically significant difference in the levels of HSP27 between healthy non-smoker and smokers without COPD (p> 0.05).

When the HSPs evaluated according to classification of airflow limitation severity; 29 (58%) COPD subjects were GOLD I-II and 21 COPD subjects (42%) were GOLD III-IV. Statistically significant difference only were found for HSP27 between

healthy non-smoker and COPD GOLD I-II (p< 0.01), healthy non-smoker and COPD GOLD III-IV (p< 0.001), smokers without COPD and COPD GOLD I-II (p< 0.05), smokers without COPD and COPD GOLD III-IV (p< 0.001), COPD GOLD I-II and COPD GOLD III-IV (p< 0.05) (Figure 1B).

There was no statistically significant difference in the IL-6 levels between groups (p> 0.05). The levels of CRP were statistically higher in COPD patients than in both healthy non-smoker and smokers without COPD (p< 0.001 for both) and the levels of MDA were significantly higher in COPD patients when compared to healthy non-smoker (p< 0.001) and smokers without COPD (p< 0.01) (Table 2).

The mean duration of disease was 6.00 ± 6.25 year, the mean PaO₂ was 63.35 ± 9.71 mmHg, PaCO₂ was 37.94 ± 5.90 mmHg, and SaO_2 was $91.27 \pm 4.40\%$, the mean 6 mwt was 368.36 ± 112.10 m and the mean symptom score was 53.47 \pm 24.51, activity score was 50.59 \pm 22.37, impact score was 38.23 ± 22.89 and total score was 44.50 ± 22.03 in COPD patients (Table 3). The mean Pa O_2 levels was significantly higher in COPD GOLD I-II patients $(68.38 \pm 7.17 \text{ mmHg})$ than COPD GOLD III-IV patients $(56.41 \pm 8.49 \text{ mmHg})$ (p< 0.001).

COPD: Chronic obstructive pulmonary disease, FEV₁: Forced expiratory volume in one second, FVC: Forced vital capacity, BMI: Body mass index. Compared with group II; a p< 0.01; c p< 0.001.

Compared with group III; $\frac{b}{b}$ p< 0.001.

Compared with group III; a p< 0.001; b p< 0.01. Results were given as median and 95% CI.

Figure 1. (A) The comparison of the serum levels of HSP27 in healthy non-smokers, smokers without COPD and COPD patients, **(B)** The comparison of serum HSP27 levels between groups when COPD evaluated according to severity of airflow obstruction in COPD.

Table 3. The duration of disease, arterial blood gases levels, 6 minute walking test and SGRQ scores in Chronic Obstructive Pulmonary Disease patients

	COPD patients $(n=50)$
Duration of disease (year)	6.00 ± 6.25
6 mwt (m)	368.36 ± 112.10
pH (mmHg)	7.41 ± 0.03
PaO, (mmHg)	63.35 ± 9.71
$PaCO$, (mm Hg)	$37.94 + 5.90$
SaO ₂ $(\%)$	$91.27 + 4.40$
SGRQ (Score)	
Symptom	53.47 ± 24.51
Activity	50.59 ± 22.37
Impact	$38.23 + 22.89$
Total	44.50 ± 22.03
COPD: Choronic obstructive pulmonary disease, SGRQ: St. Georges	
respiratory questionnaire.	

Serum levels of HSP27 showed a weak to moderate negative correlation with FEV₁, FVC and FEV₁/FVC values (Respectively, $r = -0.428$, $p < 0.01$, $r = -0.389$, $p < 0.01$, $r = -0.383$, $p < 0.01$. Only weak to moderate positive correlation were found between serum levels of HSP60 and IL-6 levels ($r = 0.327$, $p <$ 0.05). Serum levels of HSP27 showed a weak to moderate positive correlation with symptom, activity and total scores and

 $p < 0.05$ ** $p < 0.01$.

COPD: Choronic obstructive pulmonary disease, SGRQ: St. Georges respiratory questionnaire.

duration of disease (respectively, $r= 0.351$, $p< 0.05$, $r= 0.294$, p < 0.05, r = 0.316, p < 0.05). There was a weak to moderate negative correlation between HSP27 and PaO₂ ($r = -0.367$, p< 0.01). There was a weak to moderate positive correlation between HSP27 and duration of disease ($r = 0.399$, $p < 0.01$) (Table 4).

In addition, we evaluated diagnostic value of HSP27 because of increased HSP27 levels was found in COPD patients. Subjects evaluated only smokers without COPD and patients with COPD; HSP27 had an area under the curve (AUC) in the receiver operating characteristic (ROC) curve of 0.819 (0.702-0.935; 95% CI; p= 0.000). A HSP27 level of 2260 pg/ mL was taken as the cut-off between smokers without COPD and COPD patients, HSP27 had a sensitivity of 78% and specificity of 70% (ROC curve) (Figure 2A,B).

DISCUSSION

Our study shows increased levels of HSP27 in COPD patients. But the levels of HSP27 levels were not significantly different between non-smokers and smokers without COPD. Also, the levels of serum HSP27 are significantly increased in both COPD GOLD I-II and COPD GOLD III-IV patients than control subjects. When the patient's general health status was deteriorated, increased levels of HSP27 determined. The negative relationship was found between HSP27 levels and ABG and PFT parameters.

The reasons of increased release of HSPs into the extracellular environment are; the constant induction of inflammatory signals and upregulation of intracellular HSPs due to increased cellular turnover [5]. HSPs may be increased in several inflammatory disease. Elevated serum levels of HSP27 were reported in inflammatory disorders including acute coronary syndrome and chronic allograft nephropathy and increased HSP90 immunostaining was found in inflammatory regions of human atherosclerotic plaque [13- 15]. They are highly conserved chaperone proteins that regulate the folding and processing of damaged proteins and

Figure 2. (A) A cut-off level of HSP27 in smokers without COPD and COPD patients, **(B)** Receiver operating characteristic (ROC) curve indicating sensitivity and specificity of HSP27 to diagnose COPD in the Smoking study population.

therefore exert significant anti-inflammatory action and they can modulate inflammation through several mechanisms [16]. In one study, it is found that exercise modulates oxidative stress and inflammation in Aging and Cardiovascular Diseases by suppressing inflammatory pathways and also upregulates repair proteins such as HSPs [17]. Another study showed that HSP27 expression level is associated with the degree of chronic inflammation in benign prostat hypertrophy. In this study it is showed that the expression of HSP27 increased with more inflammation and this suggests that elevated inflammatory stimulation induces HSP27 expression [18]. HSP27 is also required for IL-1-induced expression of the pro-inflammatory mediators IL-6 and IL-8, and the function of HSP27 may sensitize these cells against pro-inflammatory stimuli by augmenting pro-inflammatory signaling [19]. The constant induction of inflammatory signals and increased cellular turnover result in upregulation of intracellular heat shock proteins and augmented release into the extracellular environment in COPD [5]. COPD is an inflammatory disease and the progressive inflammation in COPD continues despite cessation of smoking. For this reason, we expect that the HSPs levels increase in COPD patients. Indeed, some authors showed the increased HSPs levels in COPD patients [5,6,20].

HSP27 and HSP90 behaviors as a defensive factor against unfavorable stimuli such as heat shock and oxidative stress and it can modulate ROS and increases glutathione levels [21]. This ability results cytoprotective affects of HSP27. It is showed that HSP27 and 90 have a protective against oxidative stress [22]. The facilitator effect in the antioxidant defenses of increased HSP expression was already shown in healthy sedentary subjects [23]. HSP27 expression in smokers with or without COPD may be predominantly attributed to hypoxia and inflammation and they have protective effect in the lung cells against oxidative stress in smokers and COPD patients [20]. Increased levels of HSP27 in the lungs of smokers and especially smokers with COPD showed that increased levels of HSP27 is related with primarily oxidative stress and partly inflammation [20]. Increased serum HSP27 levels were found in subjectively healthy smokers who determined emphysema with HRCT without spirometric impairment [24]. These results shows that immune response caused by inhaled toxins in smokers' make pulmonary changes in HRCT and cause decreased HSP27 levels into the pulmonary vascular network in COPD sensitive subjects and HSP27 increases after the development of radiological COPD even thought there was no functional impairment. We found increased serum HSP27 levels in COPD patients. There was no significantly difference in HSP27 levels between healthy non-smokers and smokers without COPD subjects. Therefore we think that increased serum HSP27 levels may not be directly associated with smoking and it can only be detected increased when COPD develops. But COPD patients had higher smoking index than smokers without COPD in our study. This may affect our data. However, oxidative stress due to smoking causes the secretion of proteins but increased serum HSP27 levels in COPD patients may ascribed to other contributing factors such as hypoxia and inflammation. Also, the mean MDA levels were significantly higher in COPD patients compared with healthy non-smokers and smokers without COPD, but we did not show any relationship between HSP27 and MDA levels as an indicator of oxidant system. Further studies must be done for determining the exact antioxidant role of HSP27 in COPD patients.

Elevated HSP27 levels were reported in inflammatory disorders and HSPs expression is low under physiological conditions [14]. But HSP27 levels are temporarily increased when stress events developed and later their concentrations are decreased by termination of the acute triggering. HSP27 levels increase only when its cytoprotective effects are necessary [25]. Contrary, a continuous increase in serum HSP27 levels parallel with disease severity was shown previously [5]. Augmentation of tissue destruction in late stages of COPD and systemic inflammation of COPD may cause a systemic spillage of HSP27 into the vascular bed. Similarly, we found a continuous enhancement in serum HSP levels with severity of airflow limitation and there was an increase in HSP27 levels when respiratory function decreased and duration of disease increased. It also supports the idea that HSP27 is related with the increased tissue destruction and systemic inflammation in COPD. The relationship between serum HSP27 levels and PFT parameters as well as duration of disease interpreted that serum levels of HSP27 may be useful predictor of severity of airflow limitation in COPD stages and evaluation of response to treatment. Furthermore, we think that in addition to the systemic inflammation of COPD, hypoxia can be a contributing factor on continuous increases of HSP27, because there was a prominent hypoxia in COPD GOLD III-IV patients.

Previous experimental studies have showed increased production of HSPs in response to anoxia, presumably to help stabilize/protect protein structure/function [26,27]. Responses of HSP are organ specific [26,28]. There are a little data for the production of HSPs in the lung airway cells response to chronic hypoxia [29]. Increased HSP70 and HSP90 and unchanged HSP70 levels in lung tissue against chronic hypoxia were shown [29-31]. Consequently, the activation of heat shock response is important in stressresponsive pathways to long-term anoxic survival [32]. In our study, Pa O_2 negatively correlated with serum HSP27 levels. This interpreted that hypoxia is a prominent contrubuting factor on HSP27 levels in COPD patients.

The HSP levels are related with circulating levels of CRP and cytokines. Cytokines may increase the induction of HSPs and contrarily HSPs may decrease the release of cytokines [33]. Serum CRP and IL-6 levels were positively correlated with serum HSP levels [34]. The mechanisms of increased HSP expression due to inflammation are still not understood. In nuclear factor-IL-6 may have regulatory roles in HSP expression [35]. Different responses may be seen in HSP expression against cytokines, for example IL-6 levels increased the HSP90 levels but decreased HSP70 levels in peripheral blood mononuclear cells [36,37]. We only found a positive correlation between HSP60 and IL-6 but there was no correlation between HSPs and CRP levels. Because some of the HSP cover more than one gene, their inducible expression may be changed according to comment.

Previous studies showed that in generally increased HSP levels in COPD patients [6,20]. They may originate peripheral airways, lung interstitial cells or in other organs [5,6]. The role of extracellular HSP60 is unclear but some studies showed their pro-inflammatory effects in atherosclerosis [38,39]. Similar to Hacker's et al study, we did not find that HSP60 have a role in the pathogenesis of COPD [5]. Furthermore, we did not find any difference in serum HSP70 and HSP90 levels between the groups. The differences between study results can be due to methodological differences and differences of subject characteristics (age, gender, smoking index and respiratory function). On the other hand, HSPs are paradoxical molecules. Intracellular HSPs have beneficial and protective roles but extracellular HSPs are signal molecules for the immune system and extracellular HSPs have a modulating effect to the secretion of pro-inflammatory cytokines [40]. The exact role of serum HSPs in COPD, determines of endogenous and exogenous trigger mechanisms has to be addressed in further studies.

To our knowledge, effect of HSPs on quality of life in COPD patients has not been examined until now. Molecular chaperone expression may induce with psychological stress and psychological stress induced HSP expression was shown in rats [41,42]. For this reason, we wanted to evaluate the relationship between SGRQ scores and HSPs. SGRQ symptom, activity and total scores were significantly associated with higher serum concentrations of HSP27 in COPD patients. These findings implicate that HSP27 may be a related factor on quality of life in COPD patients. On the other hand, several factors such as duration of the disease,

disease severity and hypoxia were closely related to quality of life in COPD patients [43-45]. For this reason increased HSP27 levels may be related with decreased respiratory functions and hypoxemia on quality of life in COPD. Moreover, HSP27 may be used for the evaluation of functional status and prediction of disease severity because of HSP27 levels correlated with PFT parameters.

Proteomic analysis for determining of disease markers in COPD patients have been done previously. Serum levels of HSP27 and HSP70 may be a potential diagnostic marker and they can show disease severity [5]. Similarly these results, serum contents of HSP27 showed high sensitivity and specificity for diagnosis of COPD in our study. Because of the high sensitivity and specifity, HSP27 might be a suitable marker for diagnosis of disease according to our results.

In conclusion, the level of HSP27 was increased in COPD patients. Because smokers without COPD subjects had normal levels of HSP27 one can suppose that hypoxia is the effective factor rather than oxidant stress on serum levels of HSP27. In addition, HSP27 may be a marker of quality of life and functional status. Further investigations enrolling higher numbers of patients are needed to establish the role of HSP27 on COPD pathogenesis.

Author Contributions: Concept - R.Ü., F.D., G.K.; Desing - F.D., M.K.; Supervision - R.Ü., G.K.; Funding - S.T., D.K., R.Ü.; Materials - S.T., D.K.; Data Collection and/or Proccessing M.K., S.T., D.K.; Analysis and/or Interpretation - F.D., G.K.; Litarature Review - R.Ü.; Writer – R.Ü., F.D.; Critical Review - F.D., G.K., M.K.

Conflict of Interest: No conflict of interest was declared by the authors.

Financial Disclosure: The authors declared that this study has received no financial support.

REFERENCES

- 1. Schroder M, Kaufman RJ. ER stress and the unfolded protein response. Mutat Res 2005;569:29-63. **[CrossRef]**
- 2. Ritossa F. A new puffing pattern induced by heat shock and DNP in Drosophila. Experimentia 1962;18:571-3. **[CrossRef]**
- 3. Jammes Y, Steinberg JG, Delliaux S, et al. Chronic fatigue syndrome combines increased exercise-induced oxidative stress and reduced cytokine and Hsp responses. J Intern Med 2009;266:196-206. **[CrossRef]**
- 4. Njemini R, Abeele MV, Demanet C, et al. Age-related decrease in the inducibility of heat-shock protein 70 in human peripheral blood mononuclear cells. J Clin Immunol 2002;22:195-205.
- 5. Hacker S, Lambers C, Hoetzenecker K, et al. Elevated HSP27, HSP70 and HSP90 alpha in chronic obstructive pulmonary disease: markers for immune activation and tissue destruction. Clin Lab 2009;55:31-40. **[CrossRef]**
- 6. Cappello F, Caramori G, Campanella C, et al. Convergent sets of data from in vivo and in vitro methods point to an active role of Hsp60 in chronic obstructive pulmonary disease pathogenesis. PLoS One 2011;6:e28200. **[CrossRef]**
- 7. Global Initiative for Chronic Obstructive Lung Disease. Pocket Guide to COPD Diagnosis, Management, and Prevention. Uptadet 2015. From http://www.goldcopd.org/uploads/users/ files/GOLD_Pocket_2015_Feb18.pdf **[CrossRef]**
- 8. Cotes JE, Chinn DJ, Quanjer PH, Roca J, Yernault JC. Standardization of the measurement of transfer factor (diffusing capacity). Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society. Eur Respir J 1993;6(Suppl 16):41-52.
- 9. Jones PW, Quirk F, Baveystock C. The St. George's respiratory questionnaire. Respir Med 1991;85:25-31. **[CrossRef]**
- 10. Polatlı M, Yorgancıoğlu A, Aydemir Ö, et al. Validity and reliability of Turkish version of St. George's respiratory questionnaire. Tuberk Toraks 2013;61:81-7. **[CrossRef]**
- 11. Kuzniar T, Patkowski J. St. George's Hospital questionnaire (St. George's Respiratory Questionnaire) as an instrument for quality of life assessment in respiratory tract diseases. Pol Arch Med Wewn 2000;104:401-12. **[CrossRef]**
- 12. ATS Committee on Proficiency Standards for Clinical Pulmonary Function Laboratories. ATS statement: Guidelines for the sixminute walk test. Am J Respir Crit Care Med 2002;166:111-7. **[CrossRef]**
- 13. Park HK, Park EC, Bae SW, et al. Expression of heat shock protein 27 in human atherosclerotic plaques and increased plasma level of heat shock protein 27 in patients with acute coronary syndrome. Circulation 2006;114: 886-93. **[CrossRef]**
- 14. Djamali A, Reese S, Oberley T, et al. Heat shock protein 27 in chronic allograft nephropathy: a local stress response. Transplantation 2005;79:1645-57. **[CrossRef]**
- 15. Madrigal-Matute J, Lopez-Franco O, Blanco-Colio LM, et al. Heat shock protein 90 inhibitors attenuate inflammatory responses in atherosclerosis. Cardiovascular Research 2010; 86: 330-7. **[CrossRef]**
- 16. Sevin M, Girodon F, Garrido C, de Thonel A. HSP90 and HSP70. Implication in inflammation processes and therapeutic approaches for myeloproliferative neoplasms. Mediators Inflamm 2015;2015:970242. **[CrossRef]**
- 17. Sallam N, Laher I. Exercise modulates oxidative stress and inflammation in aging and cardiovascular diseases. Oxid Med Cell Longev 2016;2016:7239639. **[CrossRef]**
- 18. Jiang Y, Wang X, Guo Y, et al. Expression of heat shock protein 27 in benign prostatic hyperplasia with chronic inflammation. Med Sci Monit 2015;21:2976-85. **[CrossRef]**
- 19. Alford KA, Glennie S, Turrell BR, et al. Heat shock protein 27 functions in inflammatory gene expression and transforming growth factor-beta-activated kinase-1 (TAK1)-mediated signaling. J Biol Chem 2007;282:6232-41. **[CrossRef]**
- 20. Hu R, Ouyang Q, Dai A, et al. Heat shock protein 27 and cyclophilin A associate with the pathogenesis of COPD. Respirology 2011;16:983-93. **[CrossRef]**
- 21. Arrigo AP. In search of the molecular mechanism by which small stress proteins counteract apoptosis during cellular differentiation. J Cell Biochem 2005;94:241-6. **[CrossRef]**
- 22. Xanthoudakis S, Nicholson DW. Heat-shock proteins as death determinants. Nat. Cell Biol 2000;2:163-5. **[CrossRef]**
- 23. Whitam M, Fortes MB. Heat shock protein 72: release and biological significance during exercise. Front Biosci 2008;13:1328-39. **[CrossRef]**
- 24. Ankersmit JH, Nickl S, Hoeltl E, et al. Increased serum levels of HSP27 as a marker for incipient chronic obstructive pulmonary disease in young smokers. Respiration 2012;83:391-9. **[CrossRef]**
- 25. Szerafin T, Hoetzenecker K, Hacker S, et al.Heat shock proteins 27, 60, 70, 90alpha, and 20S proteasome in on-pump versus off-pump coronary artery bypass graft patients. Ann Thorac Surg 2008;85:80-7. **[CrossRef]**
- 26. Krivoruchko A, Storey KB. Regulation of the heat shock response under anoxia in the turtle, Trachemys scripta elegans. J Comp Physiol B 2010;180:403-14. **[CrossRef]**
- 27. Prentice HM, Milton SL, Scheurle D, Lutz PL. The upregulation of cognate and inducible heat shock proteins in the anoxic turtle brain. J Cereb Blood Flow Metab 2004;24:826-8. **[CrossRef]**
- 28. Hu D, Chen F, Guan C, et al. Anti-hypoxia effect of adenovirusmediated expression of heat shock protein 70 (HSP70) on primary cultured neurons. J Neurosci Res 2013;91:1174-82. **[CrossRef]**
- 29. Kim EK, Park JD, Shim SY, et al. Effect of chronic hypoxia on proliferation, apoptosis, and HSP70 expression in mouse bronchiolar epithelial cells. Physiol Res 2006;55:405-11. **[CrossRef]**
- 30. Lin HJ, Wang CT, Niu KC, et al. Hypobaric hypoxia preconditioning attenuates acute lung injury during highaltitude exposure in rats via upegulating heat-shock protein 70. Clin Sci (Lond) 2011;121:223-31. **[CrossRef]**
- 31. Konduri GG, Bakhutashvili I, Eis A, Pritchard K. Oxidant stress from uncoupled nitric oxide synthase impairs vasodilation in fetal lambs with persistent pulmonary hypertension. Am J Physiol Heart Circ Physiol 2007;292:1812-20. **[CrossRef]**
- 32. Krivoruchko A, Storey KB. Activation of the unfolded protein response during anoxia exposure in the turtle Trachemys scripta elegans. Mol Cell Biochem 2013; 374: 91-103. **[CrossRef]**
- 33. Okamura M, Takano Y, Hiramatsu N, et al. Suppression of cytokine responses by indomethacin in podocytes: a mechanism through induction of unfolded protein response. Am J Physiol Renal Physiol 2008;295:1495-503. **[CrossRef]**
- 34. Njemini R, Bautmans I, Onyema OO, et al. Circulating heat shock protein 70 in health, aging and disease. BMC Immunol 2011;12:24. **[CrossRef]**
- 35. Stephanou A, Isenberg DA, Akira S, et al. The nuclear factor interleukin-6 (NF-IL6) and signal transducer and activator of transcription-3 (STAT-3) signalling pathways co-operate to mediate the activation of the hsp90beta gene by interleukin-6 but have opposite effects on its inducibility by heat shock. Biochem J 1998;330:189-95. **[CrossRef]**
- 36. Bajramovic JJ, Bsibsi M, Geutskens SB, et al. Differential expression of stress proteins in human adult astrocytes in response to cytokines. J Neuroimmunol 2000;106:14-22. **[CrossRef]**
- 37. Stephanou A, Amin V, Isenberg DA, et al. Interleukin 6 activates heat-shock protein 90 beta gene expression. Biochem J 1997;321:103-6. **[CrossRef]**
- 38. Calderwood, SK, Mambula SS, Gray PJ, Theriault JR. Extracellular heat shock proteins in cell signaling. FEBS Lett 2007;581:3689-94.
- 39. Xu Q. Role of heat shock proteins in atherosclerosis. Arterioscler Thromb Vasc Biol 2002;22:1547-59. **[CrossRef]**
- 40. De Maio A. Extracellular heat shock proteins, cellular export vesicles and the stress observational system. A form of communication during injury, infection and cell damage. Cell Stress Chaperones 2011;16:235-49. **[CrossRef]**
- 41. Udelsman R, Blake MJ, Stagg CA, Holbrook NJ. Endocrine control of stress-induced heat shock protein 70 expression in vivo. Surgery 1994;115:611-6. **[CrossRef]**
- 42. Udelsman R, Blake MJ, Stagg CA, et al. Vascular heat shock protein expression in response to stress. Endocrine and autonomic regulation of this age-dependent response. J Clin Invest 1993;91:465-73. **[CrossRef]**
- 43. Zamzama MA, Azaba NY, El Wahsha RA, et al. Quality of life in COPD patients. Egyptian Journal of Chest Diseases and Tuberculosis 2012;4:281-9.
- 44. Halvani A, Pourfarokh N, Nasiriani K. Quality of life and related factors in patients with Chronic Obstructive Pulmonary Disease. Tanaffos 2006;5:51-6. **[CrossRef]**
- 45. Saglam M, Yagli NV, Savci S, et al. Functional capacity, physical activity, and quality of life in hypoxemic patients with chronic obstructive pulmonary disease. Int J Chron Osbtruct Pulmon Dis 2015;10:423-8. **[CrossRef]**